

## Note

**High-performance liquid chromatographic determination of cephalixin in human plasma, urine and saliva**

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Cephalexin, 7-(D- $\alpha$ -amino- $\alpha$ -phenylacetamido)-3-methyl-3-cephem-4-carboxylic acid (Fig. 1A) is one of the most commonly used oral cephalosporin antibiotics. Following oral administration, cephalexin is well absorbed and is largely eliminated by the kidney as unchanged drug [1–3]. Its elimination half-life of about an hour in normal subjects may be increased to 20–30 h in patients with severe renal dysfunction [4,5].

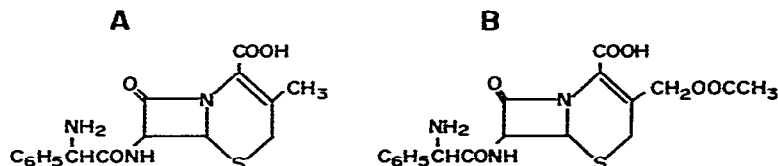


Fig. 1. Chemical structures of cephalexin (A) and cephaloglycin (B).

In severe systemic infections cephalexin dosage of 3–4 g daily for adults and 100 mg/kg/day for children have been recommended [1]. Recently, it was suggested that some pediatric patients with osteomyelitis may require as much as 200 mg/kg/day for a period of 6–16 months [6,7]. A good correlation has been observed between cephalexin serum concentration and concentration in synovial fluid of patients with suppurative arthritis [8]. Monitoring of cephalexin serum concentration is important to assure compliance and gastrointestinal absorption [9,10].

Several methods have been reported for the analysis of cephalexin using microbiological [3,11,12], spectrophotometric [13,14], fluorimetric [15–17] and polarographic [18] techniques but high-performance liquid chromatography (HPLC) is rapid, simple, sensitive and specific for cephalexin.

One HPLC method is available for determination of cephalixin in plasma [19] and two for urine [19,20]. However, no internal standard was employed placing too much reliance on accurate volume transfers and on comparing the peak of cephalixin to the standard for quantification. The procedure is unsuitable for pediatric use because a large sample (500  $\mu$ l) of plasma was required. Furthermore, these methods are not preferred for routine monitoring of plasma or urine cephalixin concentration due to long retention time for plasma (16 min) and urine (8–16 min).

Since  $pK_a$  values of cephalixin are 5.2 and 7.3 [21], and plasma protein binding is about 15% [22], cephalixin may be present in saliva. Monitoring of salivary concentration has been shown to be of value for a number of drugs [23], especially in pediatric patients because saliva can be collected by non-invasive techniques. No methods have been reported for measurement of cephalixin in saliva.

This report describes a rapid, sensitive, simple, reproducible and specific HPLC technique for the determination of cephalixin in small volume samples of plasma, urine and saliva, using cephaloglycine (Fig. 1B) as an internal standard. To demonstrate its clinical utility plasma, urinary and salivary concentrations of cephalixin in a normal subject are presented.

## MATERIALS AND METHODS

### *Chemicals and reagents*

Cephalixin, cephaloglycin and tobramycin were obtained from Eli Lilly (Indianapolis, IN, U.S.A.), carbenicillin from Roerig (New York, NY, U.S.A.), gentamicin from Sigma (St. Louis, MO, U.S.A.), and chloramphenicol and its sodium succinate ester from Parke Davis (Ann Arbor, MI, U.S.A.). Methanol (glass distilled) HPLC grade, was purchased from Mallinckrodt (Paris, KY, U.S.A.).

### *Chromatographic equipment and conditions*

The reversed-phase HPLC system consisted of Consta Metric pump II G (Laboratory Data Control, Riviera Beach, FL, U.S.A.),  $\mu$ Bondapak  $C_{18}$  column, 3.9 mm  $\times$  30 cm, 10  $\mu$ m (Waters Assoc., Milford, MA, U.S.A.), analytical fixed-wavelength UV detector, Model 153 (Beckman Instruments, Fullerton, CA, U.S.A.) and a recorder, Series 5000 (Fisher Recordall, Houston Instruments, Houston, TX, U.S.A.).

Acetic acid (0.5%) was added to methanol–water (20:80) to prepare the mobile phase. It was pumped at 2 ml/min for analysis of plasma and saliva samples and at 1.7 ml/min for urine samples. Chart speed of the recorder was set at 0.2 in./min.

### *Standards*

Cephalixin (10 mg) and cephaloglycin (10 mg) were each dissolved in distilled water (10 ml) to give a concentration of 1 mg/ml. Appropriate amounts of these standard solutions were added to the plasma, urine and saliva samples to yield concentrations of 0.2, 0.5, 1, 2, 5, 10, 20, 30, 40 and 50  $\mu$ g/ml of cephalixin. Two standard curves were constructed for cephalixin in each body

fluid: one from 0.2–10  $\mu\text{g/ml}$  using 0.5  $\mu\text{g}$  of cephaloglycin and another from 10–50  $\mu\text{g/ml}$  with 5  $\mu\text{g}$  of cephaloglycin. All samples were stored at  $-20^{\circ}\text{C}$ .

#### *Assay procedure*

Biological fluids (100  $\mu\text{l}$ ) containing known amounts of cephalixin were placed in polypropylene microcentrifuge tubes. Methanol (200  $\mu\text{l}$ ) containing internal standard was then added to the mixture. The resulting mixture was vortexed for 5 sec and then centrifuged at 9360  $g$  for 5 min. The supernatant was transferred to another set of polypropylene test tubes and evaporated at  $40^{\circ}\text{C}$  under a gentle stream of nitrogen. The residue was dissolved in 75  $\mu\text{l}$  of the mobile phase, vortex mixed for 30 sec, and 50  $\mu\text{l}$  of this were injected onto the HPLC column. The detector was set at 0.005–0.08 a.u.f.s. (wavelength 254 nm).

#### *Calculations*

The concentrations of cephalixin in the unknown plasma, urine and saliva samples were calculated by comparing its cephalixin:cephaloglycin peak height ratios with those obtained from cephalixin standard curves for plasma, urine and saliva.

#### *Recovery and precision*

Cephalixin was added to drug-free plasma, urine and saliva and then analyzed by the procedure described above but without any added internal standard. Fifty microliters of the supernatant were injected and peak heights corresponding to cephalixin measured. Absolute recoveries were calculated by comparing these peak heights with peak heights obtained by direct injection of pure standards.

Precision of the method was evaluated by analysis of plasma, urine and saliva standards containing cephalixin and cephaloglycin concentrations of 50, 25, 10 and 0.5  $\mu\text{g/ml}$ . These samples were analyzed five times by two individuals.

#### *Specificity*

Commonly used antibiotics such as gentamicin (10  $\mu\text{g/ml}$ ), tobramycin (10  $\mu\text{g/ml}$ ), carbenicillin (15  $\mu\text{g/ml}$ ), chloramphenicol sodium succinate (20  $\mu\text{g/ml}$ ) and chloramphenicol (20  $\mu\text{g/ml}$ ) were tested using this procedure for potential interference with cephalixin and cephaloglycin.

#### *Clinical application*

A normal adult volunteer (age 29) received 1 g of cephalixin as an oral suspension (Keflex<sup>®</sup>, Eli Lilly). Blood and saliva (stimulated by paraffin wax) samples were obtained simultaneously at 0, 0.25, 0.50, 1.0, 2.0, 3.0, 4.0, 6.0 h after drug administration. Urine samples were collected hourly for the first 6 h and at normal voiding hours thereafter for a total of 24 h. The specimens were stored at  $-20^{\circ}\text{C}$  and analyzed within a week.

## RESULTS AND DISCUSSION

Typical chromatograms of cephalixin and cephaloglycin in plasma, urine and saliva are shown in Fig. 2. The peaks are sharp and symmetrical allowing use of peak heights rather than peak areas to quantitate detector response. Detector response (peak height) was linear (correlation coefficients  $> 0.99$ ) over  $0.2$ – $50$   $\mu\text{g/ml}$  concentration range for cephalixin, with all curves passing through origin. Peak height ratios of cephalixin:cephaloglycin were also linear over the same concentration range.

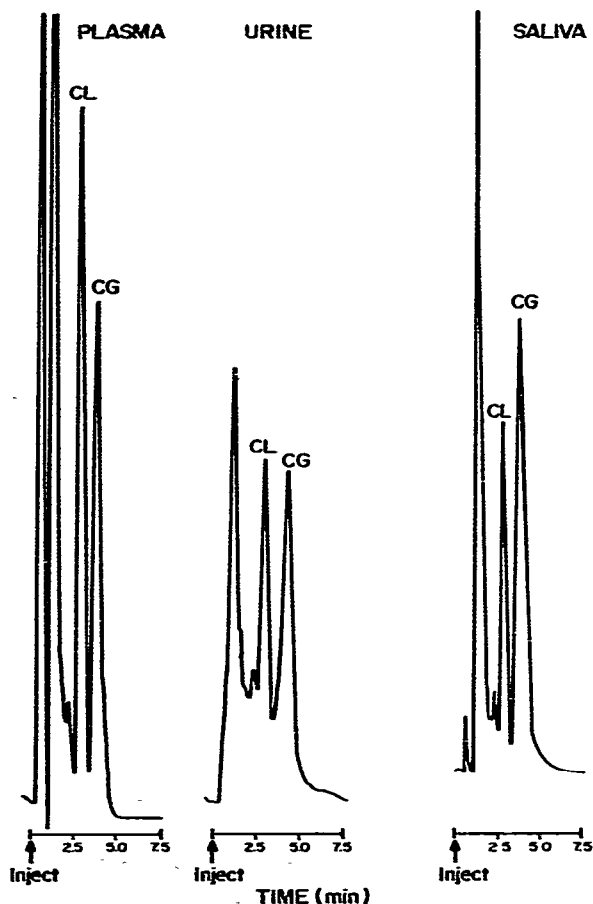


Fig. 2. Chromatograms of cephalixin (CL) and cephaloglycin (CG) in plasma (CL,  $40$   $\mu\text{g/ml}$ ; CG,  $50$   $\mu\text{g/ml}$ ), urine (CL,  $30$   $\mu\text{g/ml}$ ; CG,  $27$   $\mu\text{g/ml}$ ) and saliva (CL,  $20$   $\mu\text{g/ml}$ ; CG,  $50$   $\mu\text{g/ml}$ ). a.u.f.s.:  $0.04$ .

The retention times for cephalixin and cephaloglycin were about  $3.0$  and  $4.0$  min respectively. This is a definite advantage over the currently available HPLC procedures [19,20] because this method uses an internal standard and allows rapid analysis of cephalixin in plasma, urine and saliva.

The limit of detection was  $0.2$   $\mu\text{g/ml}$  for cephalixin in the three body fluids. Recoveries of cephalixin and cephaloglycin ranged from  $96$ – $102\%$  (Table I)

TABLE I

## RECOVERIES ON EXTRACTION OF CEPHALEXIN AND CEPHALOGLYCIN FROM PLASMA, URINE AND SALIVA

Each value is the mean of five determinations.

Compound	Concentration ( $\mu\text{g/ml}$ )	Recovery (% $\pm$ S.D.)		
		Plasma	Urine	Saliva
Cephalexin	50	101.9 $\pm$ 2.2	97.3 $\pm$ 3.3	98.1 $\pm$ 1.4
	25	97.6 $\pm$ 4.8	96.7 $\pm$ 2.7	96.5 $\pm$ 2.4
	10	96.8 $\pm$ 3.2	97.7 $\pm$ 3.8	102.0 $\pm$ 2.8
	0.5	97.1 $\pm$ 3.1	96.0 $\pm$ 4.9	97.7 $\pm$ 2.8
Cephaloglycin	50	98.1 $\pm$ 3.4	98.8 $\pm$ 2.3	98.7 $\pm$ 2.9
	25	102.0 $\pm$ 4.3	98.9 $\pm$ 3.2	100.3 $\pm$ 4.1
	10	97.8 $\pm$ 2.7	96.1 $\pm$ 4.8	97.7 $\pm$ 3.8
	0.5	96.5 $\pm$ 5.1	97.9 $\pm$ 3.7	98.1 $\pm$ 2.7

TABLE II

## REPRODUCIBILITY OF CEPHALEXIN DETERMINATION IN PLASMA, URINE AND SALIVA

In all cases  $n = 5$ .

	Concentration ( $\mu\text{g/ml}$ )	Coefficient of variation (%)		
		Plasma	Urine	Saliva
Within-day	50	2.7	3.2	2.9
	25	4.2	4.5	3.4
	10	3.0	3.9	2.3
	0.5	2.4	6.0	4.8
Day-to-day	50	2.9	4.1	4.4
	25	3.2	3.4	3.7
	10	3.0	4.9	5.0
	0.5	5.0	3.8	4.2

while precision varied from 2.3–6% (Table II). Daily variations in cephalexin concentrations were  $< 5\%$ . This assay was specific for cephalexin in that commonly used antibiotics (gentamicin, tobramycin, carbenicillin, chloramphenicol and its sodium succinate ester) did not interfere with its measurement. In these experiments cephalexin did not lose its potency during the storage for  $\leq 7$  days prior to analysis.

Plasma concentrations of cephalexin are shown in Fig. 3. The peak plasma concentration of 32  $\mu\text{g/ml}$  at 1 h after oral administration and an elimination half-life of about 1 h are consistent with reported findings [1, 24, 25]. Cephalexin concentration in saliva at 0.5 h and 1 h were 2.8 and 3.3  $\mu\text{g/ml}$ , respectively. No cephalexin was detectable beyond 1 h after drug administration. Various factors including salivary flow-rate, pH fluctuation and low partition coefficient or lipid solubility of cephalexin may be responsible for relatively low concentrations of cephalexin in saliva [23,26].

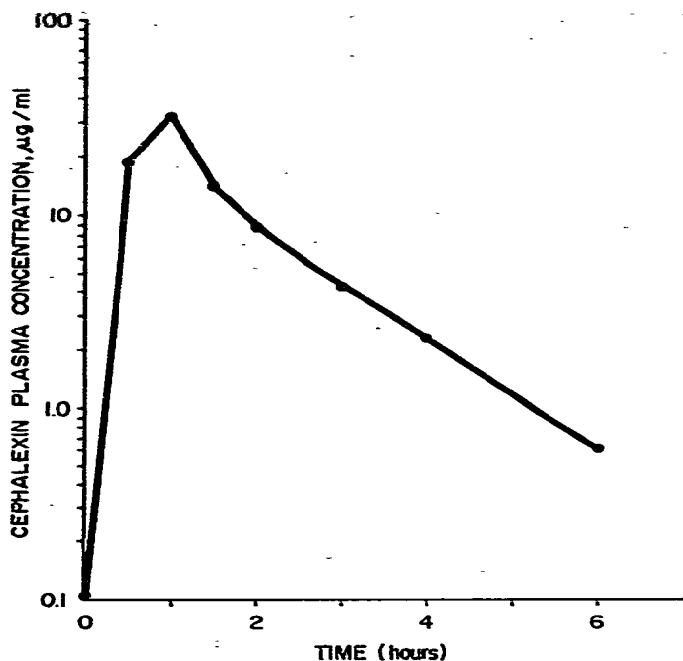


Fig. 3. Plasma concentrations of cephalixin following an 1-g oral dose.

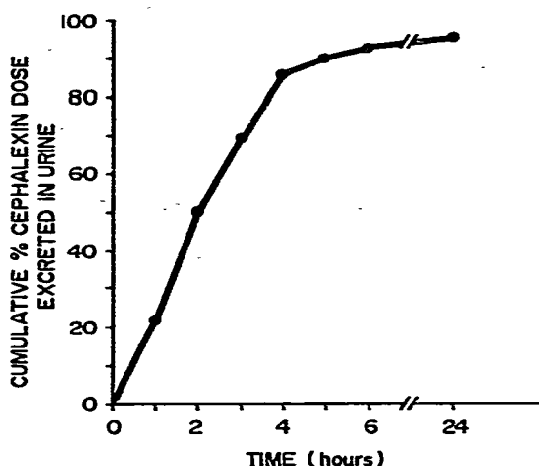


Fig. 4. Cumulative urinary excretion of cephalixin following oral administration of 1 g cephalixin.

Fig. 4 describes the cumulative urinary excretion of cephalixin during a 24-h period following drug administration. About 92% of the dose was excreted within 6 h while 95% was recovered within 24 h after cephalixin administration. These data are in agreement with other reports [1,19,20,24].

This assay system has proven simple, rapid, reproducible, sensitive and specific for determination of cephalixin in plasma, urine and saliva. The sample size required in this procedure makes it suitable for individualizing

cephalexin therapy in pediatric patients or for performing pharmacokinetic studies which requires multiple samples of biological fluids.

## REFERENCES

- 1 A. Kucers and N.M. Bennett, *The Use of Antibiotics*, J.B. Lippincott, Philadelphia, PA, 1979, pp. 225-237.
- 2 C.H. Nightingale, D.S. Greene and R.J. Quintiliani, *J. Pharm. Sci.*, 64 (1975) 12.
- 3 R.L. Perkins, H.N. Carlisle and S. Saslaw, *Amer. J. Med. Sci.*, 256 (1968) 122.
- 4 S.A. Kabins, B. Kelner, E. Waltone and E. Goldstein, *Amer. J. Med. Sci.*, 259 (1970) 133.
- 5 J.A. Linquist, J.Y. Siddiqui and I.M. Smith, *N. Engl. J. Med.*, 283 (1970) 720.
- 6 S.H. Walker, *Clin. Pediatr.*, 12 (1973) 98.
- 7 T.R. Tetzlaff, G.H. McCracken and J.D. Nelson, *J. Pediatr.*, 92 (1978) 485.
- 8 J.D. Nelson, J.B. Howard and S. Shelton, *J. Pediatr.*, 92 (1978) 131.
- 9 J.D. Nelson, *J. Pediatr.*, 92 (1978) 175.
- 10 T.R. Tetzlaff, G.H. McCracken and M.L. Thomas, *J. Pediatr.*, 92 (1978) 292.
- 11 P.E. Gower and C.H. Dash, *Brit. J. Pharmacol.*, 37 (1969) 738.
- 12 C.H. O'Collaghan, S.M. Kirby and D.R. Wishart, *Antimicrob. Ag. Chemother.*, (1968) 716.
- 13 L.P. Marrelli, in E.H. Flynn (Editor), *Cephalosporins and Penicillins*, Academic Press, New York, 1972, p. 630.
- 14 B. Casu and P. Ventura, *J. Pharm. Sci.*, 65 (1976) 211.
- 15 R. Aikawa, M. Nakano and T. Arita, *Chem. Pharm. Bull.*, 24 (1976) 2350.
- 16 A.B.C. Yu, C.H. Nightingale and D.R. Flanagan, *J. Pharm. Sci.*, 66 (1977) 213.
- 17 R.H. Barbhuiya and P. Turner, *Brit. J. Clin. Pharmacol.*, 4 (1977) 427.
- 18 E.J. Benner, *Antimicrob. Ag. Chemother.*, (1971) 201.
- 19 T. Nakagawa, J. Haginaka, K. Yamaoka and T. Uno, *J. Antibiot.*, 31 (1978) 269.
- 20 T. Nakagawa, J. Haginaka, K. Yamaoka and T. Uno, *J. Chromatogr.*, 147 (1978) 509.
- 21 M. Wirdholz, S. Budanari, L.Y. Stroumtsos and M.N. Fertig, *The Merck Index*, Merck, Rahway, NJ, 1976, pp. 1932-1933.
- 22 A.C. Kind, D.G. Kestle, H.C. Standiford and W.M.M. Kirby, *Antimicrob. Ag. Chemother.*, (1968) 361.
- 23 M. Danhof and D.D. Breimer, *Clin. Pharmacokin.*, 3 (1978) 39.
- 24 E. Finkelstein, R. Quintiliani, R. Lee, A. Bracci and C.H. Nightingale, *J. Pharm. Sci.*, 67 (1978) 1447.
- 25 H. Lode, R. Stahlmann and P. Koeppe, *Antimicrob. Ag. Chemother.*, 16 (1979) 1.
- 26 K.W. Stephen, J. McCrossan and D. Mackenzie, *Brit. J. Clin. Pharmacol.*, 9 (1980) 51.